

## SPECIFICATION

### PROCESS FOR DISCRIMINATING AND COUNTING ERYTHROBLASTS

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## BACKGROUND OF THE INVENTION

### 5 1. Field of the Invention

The present invention relates to a method for discriminating and counting erythroblasts. In particular, the present invention relates to a method for accurately discriminating and counting erythroblasts by flowcytometry.

### 10 2. Description of the Prior Art

The discrimination and counting of erythroblasts are of great use in a field of clinical examination since it provides very useful information for the diagnosis of diseases and prognosis of diseases.

15 Erythroblasts, which are also called nucleated red blood cells, are normally contained in the bone marrow, but not in peripheral blood except newborns. The appearance of erythroblasts in peripheral blood indicates a possible presence of a disease, such as leukemias, hemolytic anemia, iron deficiency anemia or pernicious anemia, and  
20 other non-hematologic/oncologic disorders. Thus the discrimination and counting of erythroblasts are very effective for the diagnosis of any such disease and for the prognosis in some of them.

Conventionally, it has been usual to prepare a smear of blood, add an appropriate dye to the smear of blood and examine it by a  
25 microscope for discriminating and counting erythroblasts. However,

such a method requires not only time-consuming, complicated pretreatment of blood for examination but also considerable expertise and skills for obtaining accurate results.

Recently, there have been available a variety of apparatuses for the fully automated discrimination and counting of leukocytes based on the principle of a flowcytometer, and there have been proposed a number of methods for analyzing blood contents using such apparatuses.

For example, Japanese Unexamined Patent Publication No. HEI 4(1992)-268453 discloses a method for discriminating and counting erythroblasts in a blood sample. This method includes the treatment of the blood sample with an acid reagent fluid of hypotonic osmolarity, the staining of nuclei of erythroblasts with a fluorescent dye solution and the detection of scattered light and fluorescent light by a flowcytometer.

Japanese Unexamined Patent Publication No. HEI 5(1993)-34251 discloses a method of determining erythroblasts. This method includes the treatment of a blood sample with an acid reagent fluid of hypotonic osmolarity, the staining of the blood sample with four kinds of dyes including Astrazon Yellow 3G and Neutral Red which are fluorescent dyes and the detection of red fluorescent light and green fluorescent light by a flowcytometer.

Published Japanese translation of PCT international publication for patent application No. HEI 8(1996)-507147 discloses a method for determining nucleated red blood cells by detecting forward

scattered light or fluorescence-side scattered light by a flowcytometer using a specific amount of a non-quaternary ammonium salt, an aliphatic aldehyde, a non-phosphate buffer, a reagent having a specific pH and a specific osmolarity and a nuclear dye such as ethidium  
5 homodimer.

U.S. Patent No. 5,559,037 discloses a method for counting erythroblasts. This method includes the lysis of erythrocytes and cell membranes of erythroblasts, the staining with a vital nuclear dye capable of staining erythroblasts but not leukocytes and the detection  
10 of scattered light at two different angles and fluorescent light by a flowcytometer.

In these methods, however, cell membranes of leukocytes as well as erythroblasts become easily damaged especially in a hematologic sample with increasing time after collection of blood.  
15 Accordingly, some leukocytes are stained with the dye that on purpose to stain erythroblasts. As a result, erythroblasts cannot be accurately determined. For example, in the case where scattered light and fluorescent light are detected, a site where erythroblasts appear overlaps or overlays a site where leukocytes appear. Especially, when  
20 lymphoid cells are damaged, it becomes more difficult to distinguish erythroblasts from damaged lymphoid cells clearly and thus the presence of erythroblasts cannot be detected accurately.

Moreover, in recent years, more and more medical laboratories send blood samples taken from patients to institutions  
25 specialized for mass examination in order to reduce costs and improve

efficiency. In such a case, it sometimes takes one day or more from the collection of blood to the examination.

Besides, it is difficult to accurately discriminate and count erythroblasts in a sample containing lymphoblasts or a sample in which chemotherapy or the like has made membranes of leukocytic cells ready to be damaged by an hemolytic agent, even if the sample does not go through change with time.

Additionally, Japanese Patent Publication No. HEI 8(1996)-1434 discloses a method for identifying nucleated red blood cells and the like. This method includes the addition of thiazole orange to a blood sample, the addition of two kinds of fluorescent labeled antibodies, anti-CD45 and anti-CD71, and the detection of signals at at least three fluorescent channels and at least two light scattering channels by a flowcytometer.

This method uses two antibodies and one fluorescent dye as reagents and has a disadvantage in that the reagents are expensive. Since the method can examine immature, nucleated red blood cells only with the combination of these specific antibodies with the specific dye, it is impossible to analyze erythroblasts cheaply by use of this method.

Further, Japanese Unexamined Patent Publication No. HEI 2(1990)-73157 discloses a method for analyzing various kinds of cells including nucleated red blood cells by detecting signals at at least three fluorescent channels and at least two light scattering channels by a flowcytometer using two kinds of fluorescent nucleic acid dyes

and a fluorescent monoclonal antibody.

According to this method, in order to discriminate erythroblasts from leukocytes, a blood sample is stained with the fluorescent monoclonal antibody, and side scattered light is measured.

5 However, since this publication lacks description about the distinction of erythroblasts from platelets and debris, it is difficult to count erythroblasts exactly by this method.

Japanese Patent No. 2620810 discloses a method for detecting fluorescent light and scattered light by a flowcytometer.  
10 The method includes the lysis of erythrocytes and the addition of a monoclonal antibody, a fixative and a nucleic acid dye which binds with DNA first.

According to this method, since a sample must be first subjected to erythrolytic treatment, the sample must be subjected to centrifugal cleaning immediately after the erythrolytic treatment.  
15 Thus absolute counting is difficult. Moreover, since this centrifugal cleaning involves complicated operation, results of detection vary significantly depending on the skill of an examiner.

Under these circumstances, there has been a demand for an  
20 easy, inexpensive method for analyzing erythroblasts accurately even in a hematologic sample collected some time before. There has also been a demand for a method for classifying and counting erythroblasts according to a degree of maturity thereof. However, such method has not been established so far.

## SUMMARY OF THE INVENTION

The present invention provides a method for discriminating and counting and staging erythroblasts comprising the steps of:

(i) staining leukocytes in a hematologic sample by adding a  
5 fluorescent labeled antibody capable of binding specifically with  
leukocytes to the hematologic sample;

(ii) raising the permeability only of cell membranes of  
erythroblasts in the hematologic sample to a nucleotide fluorescent  
dye which does not permeate a cell membrane usually;

10 (iii) staining nuclei of the erythroblasts in the hematologic  
sample with the nucleotide fluorescent dye;

(iv) subjecting the hematologic sample to flowcytometry to  
detect at least two fluorescent signals from each cell; and

(v) discriminating and counting the erythroblasts from  
15 difference in intensity between the at least two fluorescent signals.

The present invention is now described in detail by way of  
embodiments, which are not intended to limit the scope of the  
invention, with reference to the accompanying drawings in which:

## 20 BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a scattergram obtained by the method for  
discriminating and counting erythroblasts in accordance with the  
present invention;

Fig. 2 is a schematic view of Fig. 1;

25 Fig. 3 is a graph showing correlation between the ratio of

erythroblasts to leukocytes counted by the method of the present invention and that counted by a manual microscopic method;

Figs. 4A to 4C show changes with time in scattergrams obtained by the method for discriminating and counting erythroblasts in accordance with the present invention;

Figs. 5A to 5C show changes with time in scattergrams in another sample obtained by the method for discriminating and counting erythroblasts in accordance with the present invention;

Fig. 6 is a scattergram in the case where scattered light signals are combined with the method for discriminating and counting erythroblasts in accordance with the present invention;

Fig. 7 is a scattergram in which erythroblasts are distinguished from ghosts in a group of ghosts and nucleated red blood cells in Fig. 6;

Fig. 8 is a scattergram showing distribution of individual cells with intensity of green fluorescence and that of red fluorescence as coordinate axes used for NRBC staging;

Fig. 9 illustrates enclosure of each group of blood cells with windows for analysis;

Fig. 10 is a graph showing correlation between the ratio of Stage II to total erythroblast counted by the method of the present invention and that counted by a manual microscopic method;

Fig. 11 is a graph showing correlation between the ratio of State III to total erythroblast counted by the method of the present invention and that counted by a manual microscopic method.

## DETAILED DESCRIPTION OF THE INVENTION

The hematologic sample used in step (i) according to the present invention is a sample of body fluid containing leukocytes and erythroblasts, such as a sample of peripheral blood, bonemarrow or lymphoid tissue or urine, or a sample taken by apheresis.

Fluorescent labeled antibodies capable of binding specifically with leukocytes such as anti-CD45 antibody. A commercially available antibody product may be employed.

Fluorescent labeling compounds for converting the above-mentioned antibodies to the fluorescent labeled antibodies include phycoerythrin, fluorescein isothiocyanate (FITC), allophycocyanin, Texas Red, CY5, and a peridinin chlorophyll complex, which may be used singly or as a combination thereof. Preferably, these fluorescent labeling compounds have a fluorescent spectrum different from that of the fluorescent dye specific to nucleotide explained later.

Phycoerythrin and fluorescein isothiocyanate are preferred among the above compounds.

The mixture ratio of the hematologic sample to the fluorescent labeled antibody may suitably be changed depending on the state of the hematologic sample, the kind of the fluorescent labeled antibody and the like, but the mixture ratio may be about 10 : 1 to 2 : 1 by volume, for example. The mixing of the hematologic sample with the fluorescent labeled antibody may be carried out for a time period suitably set depending on temperature, but preferably at room temperature for 15 to 30 minutes or in an ice bath for 30 to 45 minutes,



for example.

In step (ii) of the present invention, the permeability only of cell membranes of erythroblasts to the nucleotide fluorescent dye is raised. Usually, the nucleotide fluorescent dye does not penetrate cell  
5 membranes.

Examples of the nucleotide fluorescent dyes are propidium iodide, N-methyl-4-(1-pyrene)vinyl-propidium iodide, ethidium bromide, TOTO-1, TOTO-3, YOYO-1, YOYO-3, BOBO-1, BOBO-3, ethidium homodimer-1 (EthD-1), ethidium homodimer-2 (EthD-2),  
10 POPO-1, POPO-3, BO-PRO-1, YO-PRO-1 and TO-PRO-1, among which propidium iodide is preferred. These nucleotide fluorescent dyes preferably have a fluorescent spectrum different from the fluorescent labeling compound for the fluorescent labeled antibody which binds specifically with leukocytes in step (i), as described above.

15 End concentration of the nucleotide fluorescent dye is about 0.003mg/L to 200mg/L, preferably about 0.03mg/L to 70mg/L, more preferably about 0.3mg/L to 35mg/L. The end concentration here means a concentration in a mixture of the hematologic sample, the fluorescent labeled antibody and the nucleotide fluorescent dye which  
20 is subjected to flowcytometry, or in the case where another agent is used as described below, in a mixture just about to be subjected to flowcytometry, preferably.

In order to raise the permeability only of the erythroblasts to the above-mentioned nucleotide fluorescent dye, for example, ① a  
25 first reagent fluid of hypotonic osmolarity containing a buffer for

maintaining pH within an acidic range is admixed to the hematologic sample, and ② admixed thereto is a second reagent fluid containing a buffer for neutralizing the mixture of the hematologic sample and the first reagent fluid in step ① and adjusting the pH of the mixture to a level suitable for staining and an osmolarity compensating agent for adjusting the osmolarity of the mixture within such a range as retains the shape and integrity of leukocytes.

The first reagent fluid in step ① is maintained at a pH within the acidic range, for example, from about 2.0 to 5.0, preferably from about 2.5 to 4.0, and more preferably from about 3.0 to 3.5, for effective lysis of erythrocytes. If the pH is too low, the reagent fluid does excessive damage to not only erythrocytes but also leukocytes, erythroblasts and the fluorescent labeled antibody specific to leukocytes. If the pH is too high, the action of the reagent fluid lysing erythrocytes is restrained.

The buffer to maintain the pH within the above-mentioned range is a buffer having an acid dissociation constant  $pK_a$  of about  $3.0 \pm 2.0$ . Examples of such buffers are malic acid and its salt, succinic acid and its salt, citric acid and its salt, phosphoric acid and its salt, and Good's buffer. The concentration of the buffer is not particularly limited so long as it is enough for maintaining the pH of the first reagent fluid at about 2.0 to 5.0. For example, the concentration may be 5 to 50 mM/l.

The first reagent fluid is required to have a hypotonic osmolarity, for example, an osmotic pressure not exceeding about 100

mOsm/kg•H<sub>2</sub>O, more preferably from about 10 to 60 mOsm/kg•H<sub>2</sub>O.

The kind of osmolarity compensating agent for adjusting the first reagent fluid within the above range of osmotic pressure is not particularly limited, but examples thereof are alkali metal salts and saccharides. For example, sodium chloride or sucrose may be used in a concentration of about 0.1g/L to 2.0g/L, for example. However, in the case where the first reagent fluid can compensate the above osmotic pressure by itself, it may not be necessary to use the osmolarity compensating agent .

The hematologic sample is treated with the first reagent fluid for a time period necessary for complete lysis of erythrocytes, for example, for about 5 to 120 seconds, preferably for about 10 to 60 seconds, more preferably for about 20 to 40 seconds.

The mixture ratio of the hematologic sample to the first reagent fluid is not particularly limited. However, in consideration of operation by a flowcytometer, the mixture ratio may be about 1 : 5 to 1 : 200 by volume, for example.

The second reagent fluid in step ② contains the buffer to neutralize the mixture of the hematologic sample and the first reagent fluid and adjust the mixture to a pH suitable for staining and the osmolarity compensating agent to adjust the mixture to an osmotic pressure suitable for retaining the shape and integrity of leukocytes.

The pH of the second reagent fluid which neutralizes acidity of the first reagent fluid and is suitable for staining is, for example, about pH5.0 to pH11.0, preferably about pH7.5 to pH10.0. The kind

of a buffer used for maintaining the pH within these ranges is not particularly limited, but a buffer having a pKa of about  $9.0 \pm 2.0$  is preferred. Examples thereof are phosphoric acid and its salt, HEPES, Tricine, and the like. The concentration of the buffer is not

5 particularly limited so far as it can maintain the second reagent fluid at a pH about 5.0 to 11.0, for example, 5 to 100 mM/l.

The osmotic pressure of the second reagent fluid suitable for retaining the shape and integrity of leukocytes is about 300 mOsm/kg•H<sub>2</sub>O to 1000 mOsm/kg•H<sub>2</sub>O, more preferably 400  
10 mOsm/Kg•H<sub>2</sub>O to 600 mOsm/kg•H<sub>2</sub>O. The kind of an osmolarity compensating agent used for adjusting the osmotic pressure within the above ranges is not particularly limited. Examples thereof are alkali metal salts and saccharides. For example, sodium chloride or sucrose may be used in a concentration of about 10.0g/L to 20.0g/L,  
15 for example.

The mixture ratio of the first reagent fluid to the second reagent fluid can be suitably adjusted depending on the pH and amount of the first reagent fluid used formerly, the concentration of the osmolarity compensating agent in the first reagent fluid, and the  
20 pH of the second reagent fluid, the concentration of the osmolarity compensating agent in the second reagent fluid and the like. For example, in the case where the pH and the osmotic pressure of the first reagent fluid are 3.0 and 16 mOsm/kg•H<sub>2</sub>O, respectively, and the pH and the osmotic pressure of the second reagent fluid are 7.5 and 400  
25 mOsm/kg•H<sub>2</sub>O, respectively, the mixture ratio of the first reagent fluid

to the second reagent fluid is preferably about 1 : 1 to 1 : 5.

For retaining the shape and integrity of leukocytes in the hematologic sample in step (ii) of the present invention, the osmotic pressure after the addition of the first reagent fluid and the second reagent fluid is preferably within the range from about 100 to 500 mOsm/kg•H<sub>2</sub>O, more preferably from 200 to 400 mOsm/kg•H<sub>2</sub>O. If the osmotic pressure after the addition of the first reagent fluid and the second reagent fluid is out of this range, it is preferable that the second reagent fluid contains an osmolarity compensating agent. The kind of the osmolarity compensating agent is not particularly limited, but is preferably a substance capable of maintaining normal biological cells at physiologic osmotic pressure, such as an alkali metal or a saccharide.

In step (iii) of the present invention, the nuclei of the erythroblasts are stained. For staining the nuclei of the erythroblasts, the hematologic sample treated in the above-described steps is stained with the nucleotide fluorescent dye. More particularly, the nucleotide fluorescent dye may be added to the first reagent fluid or second reagent fluid beforehand, and this first reagent fluid or second reagent fluid containing the nucleotide fluorescent dye may be mixed with the hematologic sample. Alternatively, a reagent fluid containing the nucleotide fluorescent dye may be separately added. The timing of adding the reagent fluid containing the nucleotide fluorescent dye is not limited, but it is preferred that the nucleotide dye is added into the first reagent fluid beforehand. A time required for the staining of the

nuclei of the erythroblasts is about 1 to 120 minutes, preferably in about 3 to 30 minutes, more preferably in about 5 to 10 minutes, after the hematologic sample is mixed with all the reagents.

The flowcytometer used in step (iv) of the present invention is not particularly limited, and may be a commercially available one. By use of such a flowcytometer, at least two fluorescent signals are detected for each cell. The fluorescent signals to be detected depend on what kind of fluorescent labeling compound is used for the fluorescent labeled antibody and what kind of nucleotide fluorescent dye is used. The fluorescent signals may be of a combination of red and green fluorescence, a combination of red and orange fluorescence, and a combination of green and orange fluorescence, among which the combination of red and green fluorescence is preferred.

In step (v) of the present invention, erythroblasts are discriminated and counted based on difference in intensity between the above-mentioned at least two fluorescent signals. For example, in the case where two fluorescent signals are detected, a two-dimensional distribution chart (scattergram) is preferably produced by plotting the fluorescence based on the fluorescent labeled antibody which binds specifically with leukocytes and the fluorescence based on the nucleotide fluorescent dye in two axes. An area in which erythroblasts appear may be defined on this two-dimensional distribution chart. The erythroblast count can be obtained by counting the number of cells in this area. An area in which leukocytes appear as well as the area in which erythroblasts appear

may be defined on this two-dimensional distribution chart. The number of cells is counted in the individual areas to obtain the erythroblast count and leukocyte count. The erythroblast count is divided by the leukocyte count and thereby the ratio of erythroblasts to leukocytes can be obtained. Alternatively a ratio between erythroblasts and total number of nucleated cell, i.e., erythroblast and leukocyte count combined, may be obtained.

Additionally, in the case where membranes of leukocytes are excessively damaged by effect of added agents and leukocytes cannot be clearly distinguished from erythroblasts in the two-dimensional distribution chart on which the nucleotide fluorescence (red fluorescence in the examples) and the fluorescence based on the fluorescent labeled antibody (green fluorescence in the examples) are plotted in the two axes, the following two steps may further be added in step (iv) for the purpose of discriminating and counting erythroblasts more accurately:

(a) Scattered light signals are also detected, and the scattered light (e.g., side scattered light signals, forward scattered light and the like, preferably side scattered light) and the fluorescence based on the fluorescent labeled antibody are plotted on two axes to produce a two-dimensional distribution chart. From this distribution chart, the group of leukocytes is identified. (see Fig. 6.)

(b) In a distribution chart on which the nucleotide fluorescence and the fluorescence based on the fluorescent labeled antibody are plotted in the two axes, an area in which the

corresponding group of leukocytes is distributed is defined. (see Fig. 7.)

(c) A boundary A between the group of leukocytes and a group of erythroblasts is marked in the two-dimensional distribution chart of (b).

Further, in the case where the end concentration of the nucleotide fluorescent dye is 0.003mg/L to 10mg/L in step (ii) , the population of erythroblasts can be classified into at least two groups according to the degree of maturity of the erythroblasts which is judged from difference basis of intensity of fluorescent light caused by the nucleotide fluorescent dye. More preferably, a suitable end concentration of the nucleotide fluorescent dye is 0.03mg/L to 3mg/L.

In this case, in step (v), it is possible to measure the maturation and count erythroblasts in different maturation stages from difference in intensity of fluorescence based on the nucleotide fluorescent dye, in other words, it is possible to determine an area where erythroblasts are present in the scattergram; determine sub-areas of groups of erythroblasts in different stages of maturity in the determined erythroblast area from difference in intensity of fluorescent light caused by the nucleotide fluorescent dye; count the number of cells in each of the sub-areas; divide the number of erythroblasts in each of the stages of maturity by the number of all the erythroblasts to obtain a ratio of erythroblasts in each of the stages of maturity to the whole erythroblasts.

The classification of erythroblasts according to their degrees



of maturity means classifying morphologic proerythroblast basophilic erythroblasts, polychromatophilic erythroblasts, orthochromatic erythroblast or the like in erythroblasts into at least two groups or into the respective groups. For example, erythroblasts are classified into proerythroblasts and three groups of basophilic erythroblasts, polychromatophilic erythroblasts and orthochromatic erythroblasts, in Example 3.

#### EXAMPLE

Examples of the method for discriminating and counting erythroblasts are now described.

##### Example 1

First, reagents having the following compositions were prepared.

Fluorescent labeled antibody:

FITC labeled anti-CD45 antibody

First reagent fluid (pH 3.0, osmotic pressure 16 mOsm/kg•H<sub>2</sub>O):

Buffering agent - Citric acid monohydrate, 2.10 g/l

- Disodium hydrogenphosphate, 0.56 g/l

Nucleotide fluorescent dye

- Propidium iodide, 100 mg/l

Purified water

Second reagent fluid (pH 7.5, osmotic pressure 420 mOsm/kg•H<sub>2</sub>O):

Buffering agent - Sodium dihydrogenphosphate dihydrate,

0.95 g/l

- Disodium hydrogenphosphate, 6.24 g/l

Osmolarity compensating agent

- Sodium chloride, 10.2g/l

Purified water

5        Fifty (50)  $\mu$ l of blood from a patient was added to peripheral  
blood to produce a hematologic sample. These bloods had been  
treated with an anticoagulant. Ten (10)  $\mu$ l of the above FITC labeled  
anti-CD45 antibody were added to the hematologic sample. This  
mixture was incubated at room temperature for about 15 minutes.  
10    Here, the blood containing erythroblasts and the peripheral blood were  
used within eight hours after they were collected.

15        Then, 500  $\mu$ l of the first reagent fluid were added to the  
mixture, which was incubated at room temperature for about 30  
seconds. To the resulting mixture, 1000  $\mu$ l of the second reagent fluid  
were added and then the mixture was incubated at room temperature  
for about 5 minutes. Then, fluorescences of wavelengths of 530nm  
(green) and 650nm (red) were detected for individual cells contained in  
the obtained hematologic sample by a flowcytometer equipped with an  
argon ion laser of 488nm wavelength as a light source.

20        Fig. 1 is a scattergram showing distribution of the cells in  
which the intensity of green fluorescence and that of red fluorescence  
are plotted in coordinate axes.

In Fig. 1, there were observed four groups: leukocytes, red  
fluorescent-stained leukocytes, erythroblasts and ghosts.

25        Analysis was made as follows: First the leukocytes and red

fluorescent-stained leukocytes were enclosed in a window (W1) as shown in Fig. 2, and all counted to obtain the leukocyte count. Next, all erythroblasts were enclosed by another window (W2) and were counted to obtain the erythroblast count. The obtained erythroblast count was divided by the obtained leukocyte count to obtain the ratio of the erythroblasts to the leukocytes.

Separately from the above example, the same hematologic sample as in Example 1 was subjected to a manual microscopic method (May Grünwald-Giemsa stain, 1,000 WBC count) discrimination and counting to obtain the morphologic erythroblast count.

Fig. 3 shows a correlation between the ratio of erythroblast determined by the flowcytometer in accordance with the present invention and that determined by the manual microscopic method.

From Fig. 3, correlation coefficient  $r$  is 0.991, which shows that the method of the present invention is remarkably accurate in discriminating and counting erythroblasts.

### Example 2

Blood containing erythroblasts from two patients other than the patient in Example 1 and peripheral blood (preserved at room temperature for 8 hours, 24 hours, 48 hours after collection) were examined about leukocytes and erythroblasts in the same manner as in Example 1. The results are shown in Figs. 4A to 4C and Figs. 5A to 5C.

The ratio of the erythroblasts to the leukocytes was also

calculated for each of the samples. The results are shown in Table 1.

Table 1

	after 8 hours	after 24 hours	after 48 hours
Sample 1	3.6%	3.6%	3.6%
Sample 2	6.1%	6.1%	6.7%

Figs. 4A to 4C and 5A to 5C and Table 1 show that the passing of time hardly affects the results of examination according to the present invention.

Example 3

Reagents having the following compositions were prepared.

Fluorescent labeled antibody:

FITC labeled anti-CD45 antibody

First reagent fluid (pH 3.0, osmotic pressure 16 mOsm/kg):

Buffering agent - Citric acid monohydrate, 2.10 g/l

- Disodium hydrogenphosphate, 0.56 g/l

Nucleotide fluorescent dye

- Propidium iodide, 1 mg/l

Purified water

Second reagent fluid (pH 7.5, osmotic pressure 420 mOsm/kg):

Buffering agent - Sodium dihydrogenphosphate

dihydrate, 0.95 g/l

- Disodium hydrogenphosphate, 6.24 g/l

Osmolarity compensating agent

- Sodium chloride, 10.2g/l

Purified water

Fifty (50)  $\mu$ l of blood from a patient was mixed with ten (10)  $\mu$ l of the above FITC labeled anti-CD45 antibody. These bloods had been treated with an anticoagulant. This mixture was incubated at room temperature for about 15 minutes.

Then, 500  $\mu$ l of the first reagent fluid were added to the mixture, which was incubated at room temperature for about 30 seconds. To the resulting mixture, 1000  $\mu$ l of the second reagent fluid were added and then the mixture was incubated at room temperature for about 5 minutes. Then, fluorescences of wavelengths of 530nm (green) and 650nm (red) were detected about individual cells contained in the obtained hematologic sample by a flowcytometer equipped with an argon ion laser of 488nm wavelength as a light source.

Fig. 8 is a scattergram showing distribution of the cells in which the intensity of green fluorescence and that of red fluorescence are plotted in coordinate axes.

In Fig. 8, there were observed six groups: leukocytes, red fluorescent-stained leukocytes, mature erythroblasts, immature erythroblasts 1, immature erythroblasts 2 and ghosts.

Analysis was made as follows: First the leukocytes and red fluorescent-stained leukocytes were enclosed in a window (W1) as shown in Fig. 9, and all counted to obtain the leukocyte count. Next, all erythroblasts were enclosed by another window (W2) and were counted to obtain the total erythroblast count. Then Stage I, Stage II

and Stage III in the window (W2) were enclosed by windows (W3), (W4) and (W5), respectively. The erythroblasts in the respective windows were counted to obtain the erythroblast staging count. The erythroblast number in the individual window was divided by the total erythroblast number to obtain the proportion of erythroblast in different stages.

Separately from the above example, the same hematologic sample as in Example 3 was subjected to a manual microscopic method (May Grünwald-Giemsa stain) discrimination and counting to classify morphologically erythroblasts into proerythroblasts, basophilic erythroblasts, polychromatophilic erythroblasts and orthochromatic erythroblasts and count the numbers thereof.

Table 2 shows results determined by the flowcytometer in accordance with the present invention and results determined by the manual microscopic method.

Table 2

Method of the Invention		Manual Microscopic Method	
Stage I	0.7%	Proerythroblast plus Basophilic erythroblast	0%
Stage II	17.6%	Polychromatophilic Erythroblasts	18%
Stage III	81.7%	Orthochromatic Erythroblast	82.0%

Table 2 shows that the results by the method the present invention agrees well with the results by the manual microscopic

method.

Example 4

Twenty-four (24) samples of bloods from patients were determined by the method of the above Example 3. Correlation between the method of the invention and the manual microscopic method was analyzed. Results were shown in Figs 10 and 11. Figs. 10 and 11 shown that the method of the invention has a definite correlation with the manual microscopic method and is capable of classifying and counting erythroblasts in the different maturity stages with great accuracy.